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3 **Variations in methods for quantification of crude ash in animal feeds**
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1
2 20 **Abstract**
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5 21 **Background:** Crude ash is categorized as an empirical method playing an important role
6 22 in the nutritional interpretation of animal feeds, allowing indirect estimation of total
7 23 organic matter.

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10 24 **Objective:** Our objective was to evaluate variations in laboratory procedures for crude
11 25 ash quantification regarding physical parameters (i.e., time, temperature) and ashing aids
12 26 and their influences on crude ash, repeatability, and discrimination power among feeds.

13
14 27 **Method:** The “control” method was based on a simple ignition time of 3 h at 550°C. The
15 28 variations are briefly described: increasing ashing time to 6 h; increasing temperature to
16 29 600°C; and using two 3-h ignition cycles at 550°C with ashing aids inclusion between
17 30 them: fresh air supply, fresh air supply plus distillate water, and fresh air supply plus
18 31 hydrogen peroxide. A color evaluation was also performed using a colorimetric
19 32 technique. Twenty four study materials from eight different feed types were evaluated.

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21 33 **Results:** The crude ash results differed among the method variations, but a consistent
22 34 decrease in the estimates was observed when liquid aids were applied, which also
23 35 improved repeatability. Ash residues did not present a consistent color pattern among
24 36 methods, but the residues were darker when the control method was applied.

25
26 37 **Conclusions:** The method of obtaining ash residues in animal feeds based on the binomial
27 38 550°C × 3 h does not have enough robustness and may overestimate crude ash in some
28 39 feeds. Adjustments in either ignition time or temperature might improve crude ash test
29 40 results, but the best test results are obtained using liquid ashing aids between two ignition
30 41 cycles.

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32 42 **Highlights:** The recommended method is based on the use of 550°C and two 3-h ignition
33 43 cycles with water added to the ash residue between cycles.

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3 44 **Introduction**
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6 45 The terms crude ash or mineral matter refer to the inorganic residue after complete
7 46 oxidation of organic matter (OM) (1, 2). However, some authors have stated that this
8 47 residue should be more properly termed as a residue on ignition (3) when oxidation is
9 48 mostly provided by burning. Generally, the ashing procedures comprise vaporization of
10 49 water and volatile compounds, and conversion of minerals into silicates, phosphates,
11 50 oxides, sulfates, and chlorides, in addition to oxidizing organic substances (2).

12
13 51 Crude ash plays an important role in the nutritional interpretation of animal feeds
14 52 by allowing indirect estimation of total OM, which encompasses all potential energy-
15 53 producing compounds (4). Moreover, ash contents are also mandatory to estimate feed
16 54 components quantified by difference, such as non-fiber carbohydrates and nitrogen-free
17 55 extract (5, 6). Crude ash estimates have been incorporated as an important input into
18 56 summative systems for the estimation of energy contents in animal diets (7-9). Actually,
19 57 any bias on the crude ash estimates might decrease the accuracy of nutritional evaluation
20 58 of animal feeds, which in turn may compromise production and culminate in economic
21 59 losses (3) due to inadequate diet formulation.

22
23 60 The standard method used to estimate % crude ash in animal feeds was first
24 61 described in the early 20th century (AOAC method 942.05; 10, 11) and is still used
25 62 worldwide as is or with some minor modifications. Briefly, a 2-g test portion is ignited at
26 63 600°C for 2 h and crude ash is reported as the residue on ignition (3).

27
28 64 Nevertheless, the crude ash in animal feeds could be categorized as an empirical
29 65 or type I method. Thus, it would be an analytical entity that determines a value that can
30 66 only be achieved in terms of the method *per se* (12). In this sense, there are no primary
31 67 reference standards that simulate the complex associations between organic and inorganic
32 68 compounds, such as those observed in feed materials. Therefore, the method itself cannot

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3 69 be validated for accuracy in determining the “true” value for the constituent. To minimize
4 70 systematic errors (i.e., bias) among laboratories, empirical methods must be followed
5 71 exactly as described in the standard manuals. Even minor variations in methodology
6 72 might result in the measurement of a different constituent (13) and compromise feed
7 73 interpretations and comparisons among feeds, laboratories, and analysts.

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9 74 The ashing procedure can be described as the submission of a test portion to a
10 75 physical binomial based on temperature and time. Despite being originally based on using
11 76 600°C, lower ignition temperatures have been suggested for the official AOAC method
12 77 since the 1940s (10, 11, 14). Excessive temperatures have been associated with systematic
13 78 bias caused by the volatilization of several minerals (3, 5, 15, 16). Thiex et al. (3) revisited
14 79 the official AOAC method 942.05 and suggested temperature adequacy down to 550°C.
15 80 Some standards for feed analysis have followed a similar pattern of temperature
16 81 adequacy, such as in ISO method 5984:2002 (17), European Commission (18), or the
17 82 latest edition of the Brazilian standards for animal feed analyses (method M-001/2; 6).

18
19 83 On the other hand, the second variable of the physical binomial applied to crude
20 84 ash quantification seems to have a more controversial pattern in the literature.
21 85 Recommendations for ashing time can range from 1 h (1) to overnight (16, 19) or until a
22 86 constant weight of residue on ignition is achieved (20). It is known that oxidation power
23 87 into the muffle furnace results from a balance between temperature and time (21). Thus,
24 88 in a logical reasoning, the higher the temperature, the shorter the ignition time, and vice-
25 89 versa. However, most methods seem to use modal times ranging from 2 to 3 hours as they
26 90 are based on temperatures between 550-600°C. In general, the use of longer ignition times
27 91 seems to be an attempt to either avoid or minimize the contamination of residue by a part
28 92 of OM that is possibly more refractory to oxidation. The influence of that refractory part

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3 93 has been associated with dark/brown colorations of the residue on ignition (3) even after
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5 94 many hours of ashing.

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7 95 In this sense, the introduction of ashing aids as a third element contributing to OM
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9 96 oxidation (i.e., besides temperature and time) has been suggested. Accordingly, Thiex et
10
11 97 al. (3) recommended a modification to the AOAC official method 942.05, which should
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13 98 encompass two 3-h ignition cycles at 550°C. The cycles are intercalated by opening the
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15 99 furnace door to guarantee a fresh air supply. In this case, the fresh air would serve as an
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17 100 oxygen supplier and would improve the oxidation of refractory organics. On the other
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19 101 hand, some authors have claimed that the dry ashing process may produce a heavy layer
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21 102 on top of the ash residue, which could hinder its complete combustion and overestimate
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23 103 ash content. In this case, a small amount of water could be poured over the ash residue
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25 104 between two ignition cycles to break up that layer (2, 3). Moreover, some variations in
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27 105 dry ashing methods have been based on the use of chemical substances as ashing aids (3,
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29 106 18, 22, 23), probably relying on the release of free radicals to speed up oxidation.

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31 107 However, despite of all the current theoretical knowledge on the dry ashing
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33 108 process, the recommendations regarding laboratory procedures are still highly variable
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35 109 (1, 2, 3, 6, 16, 18). Possibly, a “perfect” standard procedure cannot be achieved for all
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37 110 feed materials (3). Notwithstanding, the efficiency of variations in the procedures must
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39 111 be verified to assure adequate levels of method robustness while keeping an optimal
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41 112 ability to discriminate feed materials regarding their different chemical characteristics.

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43 113 Thus, our objective was to evaluate some variations in laboratory procedures for
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45 114 crude ash quantification regarding physical parameters (i.e., time, temperature) and
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47 115 ashing aids and their influences on crude ash estimates, repeatability, and discrimination
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49 116 power among feed types.

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118 **Experimental**119 *Location and study materials*

120 All analyses were performed at the Animal Nutrition Laboratory of the Animal
121 Science Department of the Universidade Federal de Viçosa, Viçosa, Minas Gerais, Brazil.

122 Eight different feed types were chosen aiming to compose a representative set
123 regarding diets offered to beef and dairy cattle: corn silage, fresh sugarcane, sugarcane
124 silage, Tifton-85 hay, soybean meal, corn grain, wheat bran, and dried distillers' grains
125 (DDG). For each feed type, three different unique materials (i.e., field replicates or
126 primary samples) were obtained from feed industries and farms located in Minas Gerais
127 State, Brazil. Therefore, our analytical set encompassed 24 study materials.

128 The high-moisture feeds (silages and fresh sugarcane) were oven-dried (55°C).
129 Then, all dry study materials were ground in a knife mill (TE-680, Tecnal, Piracicaba,
130 São Paulo, Brazil) to pass through a 1-mm screen sieve. The 24 study materials were then
131 analyzed in triplicate for dry matter (DM) content (dried overnight at 105°C, method G-
132 003/1; 6).

133 *Crude ash methods*

134 The “control” procedure herein was based on the official method of the Brazilian
135 National Institute of Science and Technology in Animal Science (INCT-CA; method M-
136 001/2; 6). Briefly:

137 (a) Weigh 2.0 g as-is of the test portion into the crucible, recording the weight of
138 crucible and test portion nearest 0.1 mg;

139 (b) Add the crucibles into the muffle furnace;

140 (c) Ignite in a furnace at 550°C for 3 h. The furnace must be adjusted to reach ignition
141 temperature within 60 min. The ignition time starts counting after ignition
142 temperature is achieved;

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3 143 (d) Allow the furnace to cool below 200°C, yet above 150°C. Then transfer crucibles
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5 144 to a desiccator; and

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7 145 (e) Cool to room temperature and weigh, recording the weight to the nearest 0.1 mg.

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9 146 For all procedures, the crucibles (5-cm diameter and 30-mL volume) were
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11 147 previously washed in running water, ashed for 3 h at 550°C, and handled and weighed as
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13 148 described above. The same digital muffle furnace was used for all procedures (Fornos
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15 149 Magnus, Belo Horizonte, Minas Gerais, Brazil).

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17 150 The following variations on the basic method were evaluated:

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19 151 1. Increased ignition temperature: all procedures were performed as previously
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21 152 described for the control, but the temperature was increased to 600°C (2, 10);

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23 153 2. Increased ashing time: all procedures were performed as previously described for
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25 154 the control, but ashing time was increased to 6 h. This specific time was chosen
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27 155 to allow a direct comparison with method variations that included ashing aids;

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29 156 3. Using fresh air as an ashing aid: the procedures were adapted from Thiex et al.
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31 157 (3). The following modifications in the control were added:

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33 158 (d) Allow the furnace to cool below 200°C and open the door to ensure a fresh
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35 159 air supply (1-2 min);

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37 160 (e) Reignite in a furnace at 550°C for 3 h. The ignition time starts counting after
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39 161 ignition temperature is achieved;

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41 162 (f) Allow the furnace to cool below 200°C, yet above 150°C. Then transfer
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43 163 crucibles to a desiccator; and

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45 164 (g) Cool to room temperature and weigh, recording the weight to the nearest 0.1
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47 165 mg;

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49 166 4. Using fresh air and water as ashing aids: the procedures were adapted from Thiex
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51 167 et al. (3) and, besides fresh air supply, it aims at breaking the superficial layer of

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3 168 residue on ignition and allowing more efficient ashing of the bottom layer in the
4 169 second ignition cycle. The modifications of procedures compared to the control
5 170 were:
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9 171 (d) Allow the furnace to cool below 100°C and open the door;
10 172 (e) Carefully add a few mL of distilled water to the residue to break it up;
11 173 (f) Reignite in a furnace at 550°C for 3 h. The ignition time starts counting after
12 174 ignition temperature is achieved;
13 175 (f) Allow the furnace to cool below 200°C, yet above 150°C. Then transfer
14 176 crucibles to a desiccator; and
15 177 (g) Cool to room temperature and weigh, recording the weight to the nearest 0.1
16 178 mg;

17 179 5. Using fresh air and hydrogen peroxide as ashing aids: the procedures were the
18 180 same as described in the latter variation, except that water was replaced by
19 181 hydrogen peroxide PA (35% or 130 vol.). In this case, besides breaking up the
20 182 upper layer, we hypothesized the hydrogen peroxide decomposition releases free
21 183 radicals, which could speed up the oxidation of the residual organic matter.

22 184 Each ashing run contained all 24 study materials. We performed three ashing runs
23 185 for each method, totaling the evaluation of 432 aliquots (i.e., test portions) (n = 3).

24 186 Following the recommendations of Thiex et al. (3), we also performed a color
25 187 evaluation of residues on ignition. Due to the small masses, the residues from the three
26 188 replicates were pooled and scored for color pattern (L* for lightness, a* for redness, and
27 189 b* for yellowness), using a Hunter MiniScan EZ colorimeter (4500 L; Hunter Associates
28 190 Laboratory Inc., Reston, Virginia, USA). Those coordinates were then converted into
29 191 RGB (i.e., red, green, and blue) coordinates using Coloroid Professional Color Plan
30 192 Designer software. After that, RGB coordinates were organized in an Excel plan in which

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3 193 each cell was filled with the correspondent solid color. For some materials, the mass of
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5 194 residues on ignition was not sufficient to allow the color evaluation.
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8 195 *Calculations and statistical analysis*
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11 196 The calculation of crude ash was performed according to the following equation:
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$$\%CA_{DM} = \frac{R - T}{W \times DM} \times 100 \quad (1),$$

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15 198 where $\%CA_{DM}$ is the crude ash as a percentage of dry matter, R is the weight of crucible
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17 199 + residue on ignition (g), T is the tare (empty) weight of crucible (g), W is the weight of
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19 200 test portion (g as-is), and DM is the dry matter content of the sample (g/g).
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22 201 The crude ash results were analyzed according to the model:
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25 202
$$Y_{ijkl} = \mu + F_i + S_{(i)j} + M_k + FM_{ik} + \varepsilon_{ijkl} \quad (2),$$

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27 203 where Y_{ijkl} is the crude ash obtained in the test portion l taken from study material j of
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29 204 feed type i and evaluated through method k, μ is the general constant, F_i is the fixed effect
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31 205 of feed type i, $S_{(i)j}$ is the random effect of study material j nested to feed i assumed to be
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33 206 NIID $(0, \sigma_{S/F}^2)$, M_k is the fixed effect of method k, FM_{ik} is the fixed effect of interaction
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35 207 between feed type i and method k, and ε_{ijkl} is the random error assumed to be NIID $(0,$
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37 208 $\sigma_{\varepsilon}^2)$.
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43 210 When necessary, means were grouped using the Fisher's multiple comparison
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46 211 procedure.
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49 212 After the first analysis of variance, data was analyzed again in an independent way
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51 213 for each method according to the model:
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54 214
$$Y_{ijk} = \mu + F_i + S_{(i)j} + \varepsilon_{ijk} \quad (3),$$

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56 215 where Y_{ijk} is the crude ash obtained in the test portion k taken from the study material j
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58 216 of feed type i, μ is the general constant, F_i is the random effect of feed type i assumed to
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3 218 be NIID $(0, \sigma_F^2)$, $S_{(i)j}$ is the random effect of study material j nested to feed type i assumed
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5 219 to be NIID $(0, \sigma_{S/F}^2)$, and ε_{ijk} is the random error assumed to be NIID $(0, \sigma_\varepsilon^2)$.
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8 220 From the adjustment of the model (3), the following relative standard deviations
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10 221 were estimated for each method variation:
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13 222 $r = \frac{\sqrt{\sigma_\varepsilon^2}}{\bar{Y}} \times 100$ (4),
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16 223 $RSD_F = \frac{\sqrt{\hat{\sigma}_F^2}}{\bar{Y}} \times 100$ (5),
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19 224 $RSD_S = \frac{\sqrt{\hat{\sigma}_{S/F}^2}}{\bar{Y}} \times 100$ (6),
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21
22 225 where r is the repeatability (%), $\hat{\sigma}_\varepsilon^2$ is the estimate of error variance $[(\% \text{ DM})^2]$, \bar{Y} is the
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24 226 average crude ash (% DM), RSD_F is the relative standard deviation among feed types
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26 227 (%), $\hat{\sigma}_F^2$ is the estimate of the variance among feed types $[(\% \text{ DM})^2]$, RSD_S is the relative
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28 228 standard deviation among study materials (%), and $\hat{\sigma}_{S/F}^2$ is the estimate of the variance
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30 229 among study materials within feed types $[(\% \text{ DM})^2]$.
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35 230 Although some reference values have been established regarding reproducibility
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37 231 (24, 25), it has been difficult to define adequate limits for repeatability. We know that the
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39 232 reproducibility evaluation cannot be performed in our work, as all procedures were
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41 233 performed in one single laboratory. However, we adopted a more functional approach to
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43 234 assess repeatability from the expected reproducibility value, which was calculated as:
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46 235 $Re = 2 \times C^{-0.15}$ (7),
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49 236 where Re is the expected reproducibility (%), and C is the average content of crude ash
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51 237 (g/g DM).
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54 238 According to Horwitz (24), repeatability should ordinarily be one-half to two-
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56 239 thirds of reproducibility. From this, we assumed that the expected conventional limits for
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58 240 repeatability could be established as:
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3 241 $L_c = 0.50 \times Re$ (8a),
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5 242 $U_c = 0.67 \times Re$ (8b),
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8 243 where L_c and U_c are the lower and upper limits for the expected conventional repeatability
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10 244 (%).

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13 245 Equation (8) is based on the assumption that reproducibility behaves exactly as
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15 246 expected (Equation 7). However, in general, a method is considered reproducible if the
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17 247 actual reproducibility falls between one-half and two-fold of expected reproducibility (24,
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19 248 25). From this, the conventional limits for repeatability may be adjusted to a tolerable
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21 249 range according to:

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23 250 $L_t = 0.50 \times 0.50 \times Re = 0.25 \times Re$ (9a),
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26 251 $U_t = 0.67 \times 2.00 \times Re = 1.34 \times Re$ (9b),
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29 252 where L_t and U_t are the lower and upper limits for the expected tolerable repeatability
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31 253 (%).

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33 254 From equations (8) and (9) we were able to understand how adequate the method
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35 255 variations behaved regarding precision/repeatability.

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37 256 All statistical evaluations were performed using the GLIMMIX procedure of SAS
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39 257 9.4. The variance components were estimated according to the restricted maximum
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41 258 likelihood method. Statistical significances were declared at $P < 0.05$.
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47 260 **Results**
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50 261 There was an interaction ($P < 0.01$) between feed types and methods on the crude
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52 262 ash. The slicing of this effect indicated that for most feed types (corn grain, DDG, wheat
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54 bran, grass hay, and fresh sugarcane) the methods did not affect ($P \geq 0.23$) the values of
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56 residues on ignition (Table 1). However, differences among methods occurred ($P < 0.01$)
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58 265 for soybean meal, corn silage, and sugarcane silage.
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3 266 The comparisons among methods within each feed type were generally different
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5 267 from each other and some overlaps were observed (Table 1). For soybean meal, the
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7 268 general pattern indicated that just increasing either time or temperature was capable to
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9 269 decrease ($P<0.05$) crude ash in comparison to the control method. However, a more
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11 270 consistent decrease compared to the control ($P<0.05$) was verified when ashing aids were
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13 271 applied. All ashing-aid variations clustered together ($P>0.05$). On the other hand, the
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15 272 mean comparisons for corn silage indicated that increasing neither temperature nor ashing
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17 273 time was sufficient ($P>0.05$) to decrease crude ash in comparison to the control. Effective
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19 274 decreases were only obtained ($P<0.05$) when liquid ashing aids were used, which did not
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21 275 differ from each other ($P>0.05$). The mean comparisons for sugarcane silage were very
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23 276 uninformative and a clear pattern could not be extracted from them.
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29 277 On average, the overall mean comparisons indicated that increasing either
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31 278 temperature or ashing time caused a consistent decrease in crude ash when compared to
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33 279 the control method ($P>0.05$, Table 1, Figure 1). In terms of ashing aids, the simple
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35 280 introduction of fresh air was not enough to decrease ($P>0.05$) crude ash in comparison to
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37 281 simply increasing the temperature or time. However, the use of liquid ashing aids caused
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39 282 an additional decrease ($P<0.05$) in crude ash compared to the other methods.
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43 283 The relative standard deviations among feed types and study materials behaved
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45 284 similarly among methods (Table 2). On the other hand, repeatability was found within
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47 285 the tolerable boundaries for all methods. However, when liquid ashing aids were added,
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49 286 the repeatability was slightly improved.
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52 287 The color evaluation was fully performed only for four feed types (Table 3). For
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54 288 the others, we faced constraints to obtain residues masses in sufficient quantities to allow
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56 289 an adequate color measurement by the colorimetric method. However, for the feed
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58 290 materials we had, there was no consistent pattern among methods. However, one specific
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3 291 point in the pattern seemed consistent enough to draw some inference. The residues on
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5 292 ignition were darker when the control method was applied, which is in line with the crude
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7 293 ash results. No systematic differences in the color pattern were verified among the other
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9 294 methods.
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14 296 **Discussion**
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17 297 The crude ash in feeds is assessed as the residue on ignition after a complete
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19 298 organic matter decomposition in a muffle furnace using a time \times temperature binomial.
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21 299 Due to its empirical nature, ashing methods have been revised (3, 26) aiming at
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23 300 minimizing their associated biases and seeking a balance between complete oxidation and
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25 301 loss of volatile minerals. In general, biases might be generated from incomplete organic
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27 matter decomposition, volatilization of some mineral compounds under specified
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29 302 temperatures, or inappropriate method application (1, 3, 16, 26).
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32 304 Despite being independent physical parameters, time and temperature do not act
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34 305 independently, and interactions between them will define the amount of crude ash
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36 306 obtained from a specific feed material (21). Once the adequate temperature is established,
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38 307 adjustments on ignition time would provide some fine-tuning on the analytical entity's
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40 308 estimates and vice-versa. On the other hand, temperatures below the minimum required
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42 309 for adequate ignition of the organic matter may not be compensated by extending ignition
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44 310 time. An equilibrium between the physical parameters of the process must be achieved to
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46 311 allow an appropriate quantification of crude ash in feeds.
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51 312 The results here confirmed that increases in either ignition time or temperature
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53 313 led, on average, to decreases in the % crude ash. It brings evidence that the physical
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55 314 binomial 550°C \times 3 h is not the best option to quantify the crude ash in feed materials.
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57 315 This was the only clear pattern pointed out by the colorimetric evaluation of residues on
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3 316 ignition. Some feed types were not sensitive to the variations in the physical parameters;
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5 317 however, others were. Thus, to achieve a more adequate robustness, the chosen method
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7 318 should focus on the feeds types that are particularly sensitive to the method.
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10 319 An ideal ignition temperature should be as low as possible to reduce volatile
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12 320 compound losses, yet high enough to ensure total carbon loss (23). A recurrent issue
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14 321 associated with excessive temperatures is the loss of minerals through volatilization (26,
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16 322 27). From this, it could be speculated that increasing the temperature to 600°C could have
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18 323 decreased ash content due to increased volatilization. However, the decreased ash content
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20 324 obtained by extending ashing time seems to allow a different explanation for that.
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22 325 Considering that the simple extension of either time or temperature led to the same
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24 326 average decrease in % crude ash, the most probable cause was an improvement in the
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26 327 elimination of some refractory organic matter, rather than an increase in volatilization.
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28 328 Once more, the inadequacy of the control procedure is evident.
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32 329 However, despite the likely improvement aforementioned, the % crude ash was,
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34 330 on average, further reduced with the use of liquid ashing aids. In this particular case, any
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36 331 loss increased by volatilization has no physical or chemical reasons to occur. However,
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38 332 the pattern obtained with the different ashing aids was not consistent among feed types.
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42 333 Despite clustering along with the liquid ashing aids for soybean meal and
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44 334 sugarcane silage, on average, the simple fresh air supply was not enough to reduce the
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46 335 crude ash at the same levels observed when liquid aids were applied. It has been stated
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48 336 that a fresh air supply between two ignition cycles could renew the oxygen supply inside
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50 337 a muffle furnace. Consequently, it could improve the release of carbon that might remain
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52 338 in the sample after the first ignition cycle (3, 28). However, such a pattern did not
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54 339 correspond to what was observed here.
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3 340 Further improvements in crude ash estimates were obtained only when liquid
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5 341 ashing aids were added between the two ignition cycles. During the ashing procedure, a
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7 342 heavy layer might be formed on the top of the residue interfering with carbon release (2).
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9 343 Then, adding liquid aids between ignition cycles might improve organic matter
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11 344 decomposition by a mechanical act, crushing the crust eventually formed in the previous
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13 345 ignition cycle and improving the degradation of refractory compounds in the second
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15 346 ignition cycle (27). Hydrogen peroxide might accelerate organic matter combustion in
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17 347 dry ashing methods (23). The decomposition of hydrogen peroxide basically occurs when
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19 348 its highly unstable oxygen-oxygen bond is broken, which can release free radicals, such
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21 349 as hydroxyl and hydroperoxyl (29). Those are highly reactive and may improve organic
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23 350 matter decomposition. However, we did not observe differences between using water and
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25 351 hydrogen peroxide as ashing aids, indicating that their effects were similar and most likely
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27 352 associated with the physical breakdown of the heavy layer in the top of the ash residue,
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29 353 allowing a better oxidation from material below the crust during the second ignition cycle.
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31 354 Considering the similarity of both liquid ashing aids, water is recommended, mainly due
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33 355 to its lower cost and ease of access.

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35 356 Several authors have considered that complete ashing is reached when the heating
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37 357 is continued until residue on ignition achieves a uniform color and is free from unburned
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39 358 particles (30, 31). In this sense, the color should be as light as possible (i.e., white, light
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41 359 gray; occasionally reddish or green). Accordingly, if the residue is dark or brown, it could
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43 360 indicate an undesirable carbon presence (3, 28, 31, 32).

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45 361 However, any visual color evaluation can be biased, as it depends on the subject's
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47 362 judgment. There is no standard color chart available for color evaluations of ash residue,
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49 363 which makes visual evaluations subjective and imprecise. Color scoring would be the
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51 364 result of a light interaction between the object and the observer's eyes, and can also vary

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3 365 according to the ambient lighting (33). That is the reason why we decided to perform a
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5 366 color evaluation using a colorimetric technique, which would avoid any subjectivity when
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7 367 scoring the residue coloration. In general, the control method (i.e, 550°C × 3 h) presented
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9 368 the darkest residues, but no clear pattern among feeds was observed for the other methods.
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12 369 Despite confirming the inadequacy of the control method, the inconsistent pattern
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14 370 among methods indicates that ash color seems to be more a characteristic of the feed
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16 371 material itself rather than a completely useful information to evaluate the ashing quality.
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18 372 Even though improvements in crude ash had been obtained using liquid ashing aids, none
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20 373 of the ash residues showed a color pattern close to white or light gray. Thus, the results
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22 374 here obtained agreed with St. John (30), who stated that neither macroscopic observations
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24 375 nor analyst's judgment for carbon presence in crude ash are useful in determining the
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26 376 optimal ashing methodology.
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31 377 Precision in our study was represented by repeatability, which is also known as
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33 378 within-laboratory variation (24). Repeatability is based on random residual variance, and
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35 379 lower values refer to a more reliable procedure for feed evaluation (34). Repeatability
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37 380 limits were proposed and calculated based on the expected reproducibility and all method
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39 381 variations exhibited repeatability within tolerable limits. However, the use of liquid
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41 382 ashing aids improved repeatability. Such a pattern reinforces our previous discussion.
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44 383 Besides overestimating crude ash, the organic matter retained in the bottom layer of ash
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46 384 residues seems to be variable among test portions. Thus, the action of the liquid ashing
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48 385 aids also improves the precision of the procedures by decreasing random variation among
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50 386 replicates.
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54 387 During the routine of feed analysis, an ideal method must also be able to allow
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56 388 adequate discrimination among and within feed types. That characteristic is expressed
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58 389 here by the relative standard deviations among feed types and study materials. Unlike
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3 390 repeatability, it is desirable that those relative standard deviations be maximized under
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5 391 any given evaluation. Hence, the capability to discriminate feeds with different
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7 392 characteristics, as well as to cluster similar feeds would be surely guaranteed. Regarding
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9 393 those requisites, all methods evaluated herein performed similarly.
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13 395 **Conclusions**

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16 396 The method of obtaining residues on ignition in animal feeds based on the
17 397 binomial $550^{\circ}\text{C} \times 3$ h does not have sufficient robustness and may overestimate crude
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19 398 ash in some feeds. Adjustments in either ignition time or temperature appear to improve
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21 399 crude ash test results, but the best results are obtained using liquid ashing aids between
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23 400 two ignition cycles. The recommended method is based on the use of 550°C and two 3 h
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25 401 ignition cycles with water added to the ash residue between cycles.
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32 33 403 **Acknowledgments**

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Table 1. Least square means for the residue on ignition (% of dry matter) in different feed types according to the crude ash method

Feed type	Ashing aid	Method ^a						SEM	P value
		Temperature (°C)	550	600	550	550	550		
		Time (h)	3	3	6	3+3	3+3		
Corn grain	-	1.18	1.18	1.17	1.13	1.15	1.13	0.689	
DDG	-	3.06	3.00	3.11	3.03	3.04	3.04	0.230	
Soybean meal	6.67a	6.49b	6.49b	6.40c	6.33c	6.32c		<0.001	
Wheat bran	6.48	6.49	6.44	6.43	6.47	6.45		0.679	
Grass hay	5.39	5.42	5.41	5.41	5.33	5.35		0.244	
Corn silage	4.71a	4.72a	4.70a	4.67a	4.59b	4.57b		<0.001	
Fresh sugarcane	2.25	2.15	2.22	2.22	2.17	2.20		0.252	
Sugarcane silage	4.20ab	4.24a	4.10c	4.18ab	4.12bc	4.11bc		0.003	
Overall	4.24a	4.21b	4.20b	4.18b	4.15c	4.15c	0.228	<0.001	

^a Means in a row followed by different letters differ at P<0.05.

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486 **Table 2. Descriptive statistics of random variabilities for the residue on ignition according to the crude ash method**

		Method						
		Temperature (°C)	550	600	550	550	550	550
		Time (h)	3	3	6	3+3	3+3	3+3
Item	Ashing aid	-	-	-	Fresh Air	Air+H ₂ O	Air+H ₂ O ₂	
Relative standard deviations (%)								
Among feed types		43.9	44.0	43.4	43.6	43.6	43.6	
Among study materials		26.5	26.4	27.1	26.6	26.6	26.5	
Repeatability (r)		2.13	2.73	2.80	2.12	1.50	1.24	
Expected reproducibility (%)		3.21	3.22	3.22	3.22	3.22	3.22	
Expected limits for r (%) ^a								
Conventional		1.61-2.15	1.61-2.16	1.61-2.16	1.61-2.16	1.61-2.16	1.61-2.16	
Maximum tolerable		0.80-4.31	0.80-4.31	0.80-4.31	0.80-4.31	0.81-4.32	0.81-4.32	

487 ^a See equations (7), (8), and (9) for more details.

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Table 3. Color pattern of the residue on ignition in different feed types according to the crude ash method

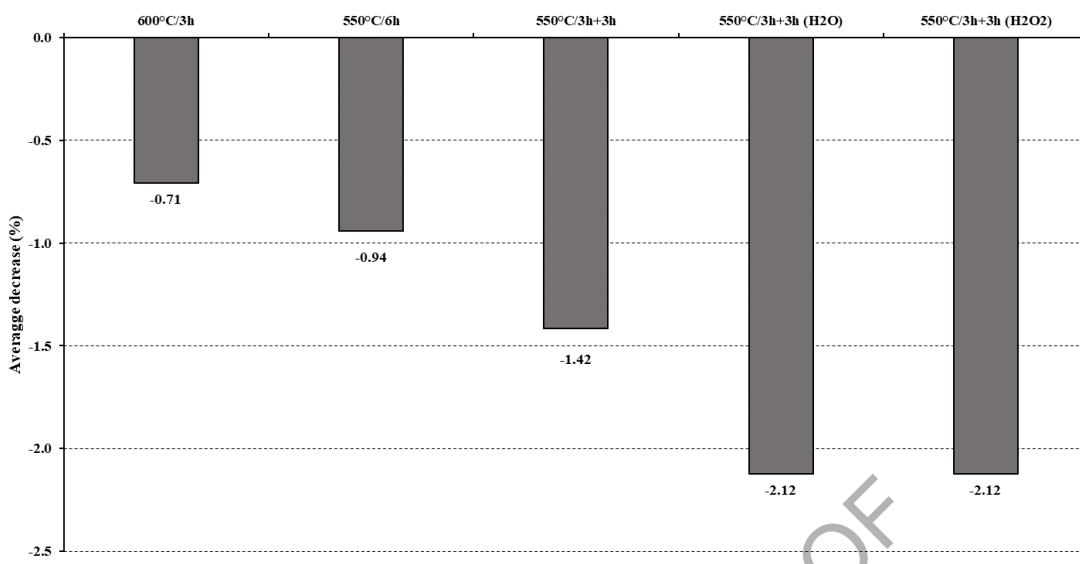
Feed type ^a	Ashing aid	Method					
		Temperature (°C)	550	600	550	550	550
		Time (h)	3	3	6	3+3	3+3
Corn grain	1	NA	NA	NA	NA	NA	NA
	2	NA	NA	NA	NA	NA	NA
	3	NA	NA	NA	NA	NA	NA
DDG	1	(160,158,156)	(156, 154, 153)	(172, 170, 168)	NA	NA	NA
	2	NA	NA	NA	NA	NA	NA
	3	NA	NA	NA	NA	NA	NA
Soybean meal	1	(164, 151, 134)	(205, 181, 154)	(185, 169, 147)	(210, 188, 159)	(213, 188, 160)	(213, 188, 160)
	2	(161, 150, 133)	(211, 185, 156)	(186, 170, 147)	(200, 178, 151)	(219, 194, 167)	(213, 188, 160)
	3	(174, 160, 141)	(216, 190, 162)	(201, 181, 155)	(215, 190, 160)	(218, 191, 165)	(211, 187, 161)
Wheat bran	1	(110, 104, 100)	(119, 117, 115)	(118, 112, 108)	(122, 177, 112)	(114, 111, 108)	(122, 118, 113)
	2	(114, 108, 103)	(120, 117, 116)	(123, 118, 113)	(128, 123, 119)	(118, 115, 113)	(151, 147, 142)
	3	(130, 125, 122)	(164, 160, 157)	(150, 146, 143)	(161, 157, 153)	(166, 159, 154)	(177, 170, 165)
Grass hay	1	(141, 134, 127)	(154, 145, 136)	(141, 133, 123)	(149, 143, 133)	(161, 150, 138)	(165, 154, 142)
	2	(145, 144, 142)	(161, 157, 152)	(152, 149, 148)	(158, 155, 154)	(157, 153, 149)	(170, 165, 160)
	3	(148, 144, 141)	(160, 154, 148)	(156, 152, 150)	(155, 151, 149)	(143, 138, 134)	(155, 149, 144)
Corn Silage	1	(125, 119, 109)	(139, 130, 118)	(133, 125, 115)	(129, 121, 110)	(145, 130, 112)	(141, 126, 109)
	2	(142, 143, 145)	(152, 151, 150)	(144, 145, 146)	(152, 152, 153)	(126, 122, 120)	(143, 140, 138)
	3	(112, 117, 122)	(127, 131, 135)	(118, 122, 128)	(117, 122, 128)	(135, 134, 135)	(119, 120, 122)
Fresh sugarcane	1	(137, 136, 135)	(142, 141, 141)	(143, 141, 141)	(144, 143, 144)	(132, 128, 126)	NA
	2	(150, 147, 145)	(162, 157, 152)	(161, 157, 153)	NA	NA	NA
	3	(152, 146, 139)	(163, 153, 140)	(149, 143, 134)	(160, 152, 142)	NA	NA
Sugarcane silage	1	(160, 155, 149)	(175, 166, 155)	(167, 161, 154)	(172, 165, 155)	NA	NA
	2	(107, 105, 105)	(122, 120, 119)	(118, 116, 116)	(108, 107, 107)	(118, 113, 109)	(122, 116, 112)
	3	(121, 113, 104)	(139, 128, 118)	(136, 126, 116)	(128, 118, 108)	(133, 118, 104)	(136, 121, 107)

490 ^aThe numbers 1 to 3 indicate the different study materials (i.e., field samples) evaluated within each feed type.

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4 491 NA: not available. The residue on ignition was insufficient for the color evaluation using the colorimeter.
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UNCORRECTED PROOF

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Figure 1. Average percentage decrease in residue on ignition according to the crude ash methods in relation to the control method (i.e., 550°C × 3 h).